

BOLD effect on calf muscle groups in elderly females with different bone mineral density

Heather T. Ma*, *Member IEEE*, James F. Griffith, Chenfei Ye, David K. Yeung, Xu Xing, Ping-Chung Leung, Jing Yuan
heather.tma@gmail.com

Abstract—This study examined the BOLD effect on calf muscles in elderly subjects with different bone mineral density. The purpose was to investigate the oxygenation characteristics in different calf muscle groups for the elderly females and compare the muscle oxygenation among groups with different bone mineral density. Temporary vascular occlusion was induced with air-cuff compression of the thigh and BOLD-MRI data curve was fitted to derive quantitative parameters. Three muscle groups, gastrocnemius muscle (lateral head), soleus muscle, and tibialis anterior muscle, were investigated individually. Quantitative CT measurement was conducted on each subject, based on which subjects were classified into normal, osteopenia, and osteoporosis groups. The BOLD signal in soleus muscle showed the lowest minimum ischemic value during ischemia and the steepest slope during hyperemia. As soleus muscle is mainly composed by slow-twitch oxidative muscle fibers, current results may be due to a higher vascular bed density and better endothelial function in such muscle. By t-test, the half-life of the BOLD signal decay during ischemia in both gastrocnemius and soleus muscles was significantly prolonged in osteoporosis group, indicating a degenerated muscular oxygen metabolic capacity in osteoporotic patients.

I. INTRODUCTION

Lower-extremity peripheral artery disease affects roughly 1 in 20 adults aged above 55 years [1]. Current methods used to measure skeletal muscle blood flow are either limited by spatial resolution (e.g. PET) or are invasive (e.g. DCE-MRI). Development of non-invasive techniques for evaluating vascular function would be highly desirable. Functional MRI relies on the blood oxygenation level-dependent (BOLD)

Research supported by the Basic Research Foundation (Outstanding Young Investigator Track) of Shenzhen (JC201005260124A), Research Grants Council of the Hong Kong Special Administrative Region, China (Project No. 465111), High-end Talent Oversea Returnees Foundation of Shenzhen (KQC201109020052A), and National Natural Science Foundation of China (81000647).

Heather T. Ma, Chenfei Ye, and Xu Xing are with the Department of Electronic & Information Engineering, Harbin Institute of Technology Shenzhen Graduate School, Shenzhen, China. (Corresponding author: Heather T. Ma. Phone: +86-755-26033608; fax: +86-755-26033608; e-mail: heather.tma@gmail.com).

James F. Griffith and David K. Yeung are with the Department of Diagnostic Radiology & Organ Imaging, The Chinese University of Hong Kong, Hong Kong, China. (email: griffith@cuhk.edu.hk; dkeyeung@cuhk.edu.hk)

Ping-Chung Leung is with the Jockey Club Centre for Osteoporosis Care and Control, The Chinese University of Hong Kong, Hong Kong, China. (email: pingcleung@cuhk.edu.hk)

Jing Yuan is with the Medical Physics and Research Department, Hong Kong Sanatorium & Hospital, Happy Valley, Hong Kong, China.

effect that is sensitive to changes in local tissue deoxyhemoglobin concentration [2]. This technique is widely used in human brain activation mapping and also in skeletal muscle perfusion studies [3-7]. Skeletal muscle oxygenation level varies with local metabolic process and oxygen supply by the feeding arteries or capillaries. Muscle BOLD effect in the lower limb may be observed by applying a T2*-weighted sequence while an air-cuff is inflated above the knee to induce ischemia. On releasing air-pressure, post-ischemic reactive hyperemia may be assessed by measuring the rate of T2* signal recovery [6-10].

The BOLD effect in the skeleton muscle can reflect hemoglobin oxygenation inside the muscle's capillary bed, which is mainly influenced by blood inflow, fluid shifts, metabolic factors, vascular architecture, and magnetic field angulation [1, 11]. Previous studies indicated that osteoporotic bone had a reduced perfusion in bone marrow while remained a normal perfusion function in the surrounding muscles [12,13]. Perfusion process reflects the blood inflow in the tissue, which only partially indicates the local oxygenation situation. If the muscle oxygenation keeps at the normal level in osteoporotic patients is unknown yet. BOLD examination on muscle would reveal the muscle function change in osteoporosis in terms of the muscle oxygenation function.

Therefore, this study was proposed to examine the BOLD effect on calf muscles in elderly subjects with different bone mineral density (BMD) to investigate the oxygenation characteristics in different calf muscle groups and the difference in osteoporotic patients. Temporary vascular occlusion was induced with air-cuff compression of the thigh and BOLD-MRI data curve was fitted to derive quantitative parameters. Three muscle groups, gastrocnemius muscle (lateral head), soleus muscle, and tibialis anterior muscle, were investigated individually. Subjects were classified into three groups (normal, osteopenia, and osteoporosis) based on the bone mineral density (BMD) measured by the quantitative CT (QCT).

II. METHODOLOGY

A. Subjects

Subjects were recruited in a common community by public advertisement. In order to avoid gender influence, only female subjects were included in current study. Subjects were excluded if they had (a) clinical or imaging evidence of renal osteodystrophy or other metabolic bone disease other than osteoporosis or a known malignancy, (b) a history of lumbar spinal surgery or irradiation, or (c) MR imaging evidence of large intravertebral disk herniation, hemangioma, or moderate to severe vertebral fracture of L3. Finally, 87 subjects (age 65.4

±4.3yrs) in total were involved in this study. The study was approved by the Ethics committee, Chinese University of Hong Kong with all participating subjects submitting written consent.

B. Data acquisition

Quantitative CT (QCT) was conducted on subjects' L3 of lumbar spine by Lightspeed VCT 64. Standard phantom was put under the lumbar during the imaging. Bone mineral density was then derived according to the CT values from the bone and the phantom.

Each subject was also required to undergo BOLD imaging of the lower limb using a 3T whole-body scanner (Achieva TX, Philips Healthcare). An eight-channel SENSE knee coil was used for signal reception and a T2*-weighted sequence (TR/TE 372/40 ms; slice thickness 5 mm; NEX 1; FOV 330 mm; dynamic measurements 2400; scan time 900s) was employed. An air-cuff was placed just above the left knee with an imaging plane selected 20-25 cm away from the distal side of the air-cuff. Air-cuff pressure applied was 50mmHg above systolic pressure. The cuff was inflated at 1 min and maintained for 5 minutes. Followed cuff deflation, the scanning continued for a further 9 minutes. Regions of interest (ROI) were drawn on the gastrocnemius muscle (lateral head), soleus muscle, and tibialis anterior muscle on the T1 images (Fig.1), which was co-registered onto the BOLD image series. The BOLD signals in the corresponding ROIs were measured from T2* images.

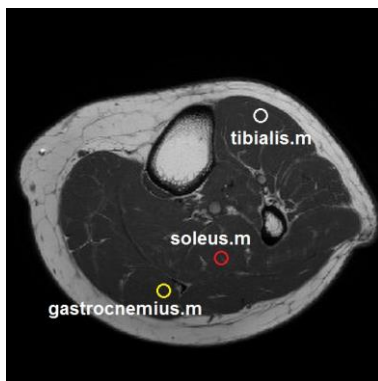


Fig 1. T1 image of the calf slice showing the location of ROIs drawn in gastrocnemius muscle (yellow circle), soleus muscle (red circle), and tibialis muscle (white circle).

C. Signal processing and data analysis

The BOLD-MRI signal was first normalized by the resting stage signal intensity, which is the starting period of the imaging. A curve fitting model was employed to analyze BOLD-MRI signals using PRISM software (ver. 5). After signal normalization, four parameters were selected to characterize the T2* time course as shown in Fig 2:

1. **MIV**: the minimum ischemic value, which is the minimal BOLD signal during the ischemic phase relative to baseline;
2. **HPV**: hyperemia peak value, which refers to the maximum height of T2* signal during hyperemia relative to baseline.

3. **Half-life**: the time interval for the BOLD signal decay to half MIV during ischemia as measured from the beginning of the ischemia;
4. **Slope**: the ratio between the HPV+baseline and the time interval for hyperemia, which is a measure to indicate the rate of T2* signal surge at the moment of cuff release;

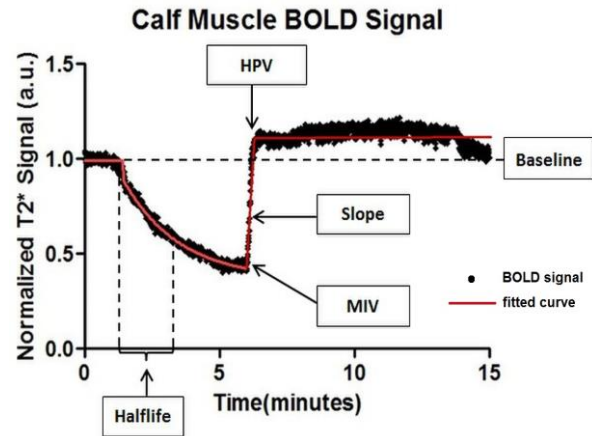


Fig 2. An example of curve fitting model for normalized T2* signal

Subjects were classified into three groups based on the BMD values derived from QCT measurement. According to the American College of Radiology (ACR) [17]: a BMD $\leq 80 \text{ mg/cm}^3$ indicates osteoporosis; a BMD $\leq 120 \text{ mg/cm}^3$ and $> 80 \text{ mg/cm}^3$ indicates osteopenia; and a BMD above 120 mg/cm^3 is considered normal.

ANOVA and t-test were performed for the key parameters to distinct difference among muscle groups and BMD groups. All statistical analyses were performed using SPSS 16 and $p=0.05$ was set as the statistically significant level.

III. RESULTS

The ANOVA of the parameter values derived from the three muscle groups for the 87 subjects are shown in Table 1. Significant differences in Slope and MIV were observed among the muscle groups ($p < 0.05$). Soleus muscle has the highest Slope but the smallest MIV value, compared to the other two muscle groups. Therefore, the soleus muscle should have a steeper enhancement phase during the hyperemia phase. The comparison is further elucidated by Fig.3.

Table 1: Muscle groups comparison by ANOVA

Muscle Groups	Slope (a.u./min)	Half-life (min)	HPV (%)	MIV (%)
Gastroc	1.5±0.6	3.0±2.5	3.9±8.6	62.3±14.3
Soleus	1.8±0.7	4.6±4.8	5.8±8.6	55.9±13.0
Tibialis	1.3±0.4	2.3±2.0	5.5±7.4	68.2±11.0
P-value	<0.0001	0.065	0.243	<0.0001

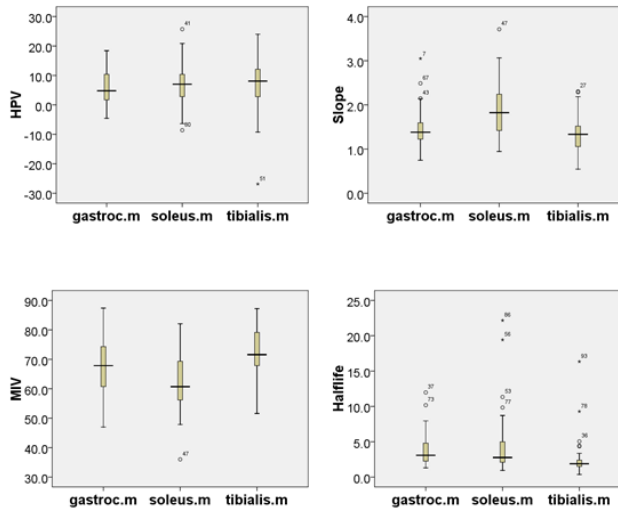


Fig 3. Comparison of the four parameters (Slope, Half-life, MIV, HPV) among 3 muscle groups.

The comparison of the muscle parameters among different BMD groups is summarized by Table 2.

Table2: ANOVA for muscle parameter among BMD groups

Groups		gas.m	soleus.m	tibia.m
Half-life (min)	0	3.3±2.5	2.9±1.6	2.2±0.8
	1	2.3±1.5	3.2±3.7	2.2±1.6
	2	4.3±3.7	4.5±6.1	2.9±3.4
<i>P-value</i>		--	--	--
MIV (%)	0	61.5±16.8	56.7±12.5	69.1±11.0
	1	62.9±14.0	56.7±12.5	67.4±11.7
	2	60.7±12.4	58.3±11.8	68.5±9.7
<i>P-value</i>		0.912	0.447	0.902
slope (a.u./min)	0	1.6±0.6	1.8±0.7	1.4±0.6
	1	1.5±0.5	2.0±0.7	1.3±0.4
	2	1.4±0.6	1.7±0.5	1.2±0.3
<i>P-value</i>		0.601	0.206	0.269
HPV (%)	0	3.6±10.4	3.5±9.0	5.7±5.0
	1	3.6±10.4	6.7±8.8	5.3±8.6
	2	2.1±9.7	5.7±7.8	6.0±6.9
<i>P-value</i>		0.531	0.388	0.939

Note: 0 represents normal group; 1 represents osteopenia group, and 2 represents osteoporosis group

No significant change by ANOVA is observed from Table 2. However, the Half-life mean value of both gastrocnemius and soleus muscles is much higher in osteoporosis group compared to the normal group. Due to the large variance, ANOVA was failed in the analysis. When we further investigated the difference by t-test, the Half-life value was significantly larger in osteoporosis group compared to the normal group. This result indicates that osteoporotic patients would have a longer decay half-life during the ischemia stage for the gastrocnemius and soleus muscles.

Table3: p-value from the t-test on Half-life among BMD groups

Muscle	0 vs 1	1 vs 2	0 vs 2
gastroc.m	0.045	0.36	0.04
soleus.m	0.06	0.13	0.016

Note: 0 represents normal group; 1 represents osteopenia group, and 2 represents osteoporosis group

IV. DISCUSSIONS

Functional MRI is a promising tool for noninvasive investigation of ischemia and reperfusion process of skeletal muscle. Although no standard procedure for experiment and data, many studies have characterized the BOLD signal intensity curve by quantitative parameters [1]. BOLD signal changes reflect muscle oxygenation and in particular the degree of hemoglobin oxygenation. Several physiologic parameters have been postulated to influence the muscle BOLD signal, such as muscle blood volume, perfusion, fluid shifts, metabolic factors, vascular architecture, and magnetic field angulation [6,9]. Due to their mutual interference and paradigm-dependence, dissecting the impact of every single factor on the muscle BOLD effect is difficult. This study was proposed to investigate the muscle oxygenation difference in the elderly females with different BMD by using quantitative parameters, such as MIV, HPV, Slope, and Half-life.

First, the BOLD effect of the three muscle groups was investigated for all the subjects. It was found that soleus muscle had a significant lower MIV during ischemia than the other calf muscles. Parameter MIV reflects oxygen utilization while Slope reflects vascular reactivity [1]. Conversely, the soleus also showed the steepest slope upon reperfusion indicating highest vascular reactivity or density. These changes are in line with expected findings. The soleus muscle mainly helps maintain balance and is mainly consists of slow-twitch oxidative muscle fibers which tend to have a higher capillary density and myoglobin content compared to fast-twitch dominant muscles, such as the gastrocnemius muscle. In this study, using a non-invasive technique, we have been able to demonstrate different oxygen requirements, metabolism and re-perfusion capability in different calf muscle groups. Vascular compromise could potentially affect some muscle groups differently to others.

Secondly, a significantly prolonged Half-life was observed in gastrocnemius and soleus muscles of osteoporotic subjects compared to those with normal BMD. Parameter Half-life reflects the rate of oxygen metabolism or the efficiency of the oxygen uptake. This difference indicates a reduction of the muscular oxygen metabolic capacity in osteoporotic patients. Taking the comparison results of the muscle groups, it appears that muscle group with more slow-twitch fibers would have a prolonged Half-life during ischemia. The current result may imply that in osteoporotic patients, more muscle fibers transformed to slow-twitched fibers. Actually this implication can be supported by a previous study on muscle fiber of osteoporotic patients by muscle biopsy [14]. In that study, fast-twitched muscle fiber was found positively correlated to the BMD. In other words, osteoporotic patients would have more slow-twitched muscle fibers compared to normal subjects. Such muscle degeneration

in osteoporosis should be further investigated, which may reveal other mechanism of osteoporosis.

ACKNOWLEDGMENT

This study is supported by the Research Grants Council of the Hong Kong Special Administrative Region, China (Project No. 465111), Basic Research Foundation (Outstanding Young Investigator Track) of Shenzhen (JC201005260124A), High-end Talent Oversea Returnees Foundation of Shenzhen (KQC201109020052A), and National Natural Science Foundation of China (81000647).

REFERENCES

- [1] B. Jacobi, G. Bongartz, et al, "Skeletal muscle BOLD MRI: from underlying physiological concepts to its usefulness in clinical conditions," *J Magn Reson Imaging*, 35:1253-1265, 2012.
- [2] Ogawa S, Lee TM, Kay AR, Tank DW, "Brain magnetic resonance imaging with contrast dependent on blood oxygenation," *Proc Natl Acad Sci U S A*, 1990 Dec, 87(24):9868-72.
- [3] A.C. Schulte, M. Aschwanden, D. Bilecen, "Calf muscles at blood oxygen level-dependent MR imaging aging effect at postocclusive reactive hyperemia," *Radiology*, 247(2): 482-489, 2008.
- [4] Lebon V, Brillault-Salvat C, Bloch G, Leroy-Willig A, Carlier PG, "Evidence of muscle BOLD effect revealed by simultaneous interleaved gradient-echo NMRI and myoglobin NMRS during leg ischemia," *Magn Reson Med*, 1998, 40:551 - 558.
- [5] Donahue KM, Van Kylene J, Guven S, et al, "Simultaneous gradient-echo/spin-echo EPI of graded ischemia in human skeletal muscle," *J Magn Reson Imaging*, 1998, 8:1106 - 1113.
- [6] Noseworthy MD, Bulte DP, Alfonsi J, "BOLD magnetic resonance imaging of skeletal muscle," *Semin Musculoskelet Radiol* 2003, 7:307 - 315.
- [7] Thompson RB, Aviles RJ, Faranesh AZ, et al, "Measurement of skeletal muscle perfusion during post-ischemic reactive hyperemia using contrast-enhanced MRI with a step-input function," *Magn Reson Med*, 2005;54:289 - 298.
- [8] J. F. Tousaint, K. K. Kwong, F. O. Mkpuru, et al, "Perfusion changes in human skeletal muscle during reactive hyperemia measured by echo-planar imaging," *Magn Reson Med*, 1996;35:62 - 69.
- [9] Lederemann HP, Heidecker HG, Schulte AC, et al, "Calf muscles imaged at BOLD MR: correlation with TcPO₂ and flowmetry measurements during ischemia and reactive hyperemia—initial experience," *Radiology*, 2006;241:477 - 484.
- [10] Schulte AC, Aschwanden M, Bilecen D, "Calf muscles at blood oxygen level-dependent MR imaging: aging effects at postocclusive reactive hyperemia," *Radiology*, 2008, 247:482 - 489.
- [11] Towse TF, Slade JM, Ambrose JA, Delano MC, Meyer RA, "Quantitative analysis of the post-contractile blood-oxygenation-level-dependent (BOLD) effect in skeletal muscle," *J Appl Physiol*, 2011, 111:27-39.
- [12] J. F. Griffith, D. K. W. Yeung, G. E. Antonio, S. Y. S. Wong, T. C. Y. Kwok, J. Woo and P. C. Leung, "Vertebral Marrow Fat Content and Diffusion and Perfusion Indexes in Women with Varying Bone Density: MR Evaluation," *Radiology*, vol. 241, no. 3, pp. 831-838, 2006.
- [13] Xinxin Zhao, Heather T. Ma, Jun Zhu, Xu Xing, James F. Griffith, Ping-Chung Leung, "Bone Marrow Perfusion of Females with Varied Bone Mineral Density: a Study by Muscle-based Model", Proceedings of IEEE-TENCON 2013, Xi'an, China, Oct 2013.
- [14] Wing-Hoi Cheung, Wing-Sze Lee, Ling Qin, et al, "Type IIB human skeletal muscle fibers positively correlate with bone mineral density irrespective to age," *Chinese Medical Journal*, 2010, 123(21): 3009 - 3014.
- [15] Utz W, Jordan J, Niendorf T, et al, "Blood oxygen level-dependent MRI of tissue oxygenation: relation to endothelium-dependent and endothelium-independent blood flow changes," *Arterioscler Thromb Vasc Biol*, 2005, 25:1408 - 1413.

- [16] Lederemann HP, Schulte AC, Heidecker HG, et al, "Blood oxygenation level-dependent magnetic resonance imaging of the skeletal muscle in patients with peripheral arterial occlusive disease," *Circulation* 2006, 113:2929 - 2935.
- [17] Klarh € ofer M, Mad € orin P, Bilecen D, Scheffler K, "Assessment of muscle oxygenation with balanced SSFP: a quantitative signal analysis," *J Magn Reson Imaging*, 2008, 27:1169 - 1174.
- [18] Kos S, Klarh € ofer M, Aschwanden M, Scheffler K, Jacob AL, Bilecen D, "Simultaneous dynamic blood oxygen level-dependent magnetic resonance imaging of foot and calf muscles: aging effects at ischemia and postocclusive hyperemia in healthy volunteers," *Invest Radiol*, 2009, 44:741 - 747.
- [19] American College of Radiology. ACR practice guideline for the performance of quantitative computed tomography (QCT) bone densitometry[J]. 2008.
- [20] Huegli RW, Schulte AC, Aschwanden M, Thalhammer C, Kos S, Jacob AL, Bilecen D, "Effects of percutaneous transluminal angioplasty on muscle BOLD-MRI in patients with peripheral arterial occlusive disease: preliminary results," *Eur Radiol*, 2009, 19:509 - 515.
- [21] Langham MC, Floyd TF, Mohler ER 3rd, Magland JF, Wehrli FW, "Evaluation of cuff-induced ischemia in the lower extremity by magnetic resonance oximetry," *J Am Coll Cardiol*, 2010, 55:598 - 606.
- [22] Sanchez OA, Copenhaver EA, Elder CP, Damon BM, "Absence of a significant extravascular contribution to the skeletal muscle BOLD effect at 3 T," *Magn Reson Med*, 2010, 64:527 - 535.
- [23] Raj D, Paley DP, Anderson AW, Kennan RP, Gore JC, "A model for susceptibility artefacts from respiration in functional echo-planar magnetic resonance imaging," *Phys Med Biol*, 2000, 45:3809 - 3820.
- [24] Barry RL, Menon RS, "Modeling and suppression of respiration-related physiological noise in echo-planar functional magnetic resonance imaging using global and one-dimensional navigator echo correction," *Magn Reson Med*, 2005, 54:411 - 418.
- [25] Cai J, Wang L, Wu P, Li Z, Tong L, Sun S, "Study on oxygen enrichment from air by application of the gradient magnetic field," *J Magn Magn Mater*, 2008, 320:171 - 181.
- [26] Duteil S, Wary C, Raynaud JS, et al, "Influence of vascular filling and perfusion on BOLD contrast during reactive hyperemia in human skeletal muscle," *Magn Reson Med*, 2006, 55:450 - 454.
- [27] Versluis B, Backes WH, van Eupen MG, et al, "Magnetic resonance imaging in peripheral arterial disease: reproducibility of the assessment of morphological and functional vascular status," *Invest Radiol*, 2011, 46:11 - 24.